

Developments and challenges in the detection of doping with peptide hormones and related substances

New techniques for extraction, separation, and detection of EPO

Dr. Zhen Liu

State Key Lab of Analytical Chemistry for Life Science Nanjing University

Rome

June 14-16, 2011

Outline

- Ø Nanoparticles-based extraction-capillary zone electrophoresis-native fluorescence detection of EPO
- Ø Boronate affinity monolithic capillaries for specific extraction of EPO

Outline

- Ø Nanoparticles-based extraction-capillary zone electrophoresis-native fluorescence detection of EPO
- Ø Boronate affinity monolithic capillaries for specific extraction of EPO

The IOC-validated EPO assay



Our proposed strategy



Capillary zone electrophoresis (CZE)-UV absorbance detection of EPO standard



Capillary, bare fused-silica capillary with 60 cm total length (50 cm effective length) and 50 μ m id; capillary temperature, 35 °C; separation voltage, 15 kV; BGE, 10 mM sodium acetate buffer containing 7 M urea, 10 mM Tricine, 3.9 mM putrescine, and 100 mM NaCl at pH 5.50; UV detection wavelength, 214 nm; sample, 1 mg/mL EPO, injected at 0.5 psi for 5 s.

Capillary zone electrophoresis (CZE) with online sample concentration of EPO standard



Principle of large volume sample stacking-reversed pH junction (LVSS-RPHJ)



CZE with LVSS-RPHJ of EPO

Sample and injected zone length (A) 0.02 mg/mL, 24.5 cm; (B) 0.01 mg/mL, 49 cm.

The detection sensitivity can be improved by 50–100 times, but at the price of resolution loss.

In-lab built deep UV LIF detection (dual channel LIF)



Beckman Coulter PACE MDQ system



Fiber and fiber adapter



Comparison of the LIF detection of intact protein and fluorescent dye labeled protein



Native fluorescence

More sensitive as compared with UV absorbance (1 order of magnitude higher)

Original properties are preserved

Label-free

Fluorescent labeling

Much more sensitive as compared with native fluorescence (50 folds higher) Variation of the original properties Multiple labeling

Deep UV LIF detection of EPO and comparison with UV absorbance



Label-free, identical finger-print profile

Improved detection sensitivity, 1 order of magnitude higher as compared with UV absorbance

CZE-LINF with on-line sample concentration of EPO standard



CZE with LVSS-RPHJ of EPO

Sample concentration: 0.001 mg/mL (3×10^{-8} mol/L).

Procedure of magnetic particles-based extraction



Amino-functionalized magnetic nanoparticles (Amino-MNPs)





Random immobilization of antibodies through glutaraldehyde coupling



Extraction of AFP by anti-AFP immobilized MNPs



Oriented immobilization of anti-AFP IgG after oxidation of the carbohydrate moiety



Extraction of AFP by anti-AFP immobilized MNPs



Extraction of EPO by anti-EPO immobilized MNPs



Oriented immobilization of Anti-EPO IgG to protein A-immobilized magnetic beads



Optimization of experimental conditions (1)







Optimization of experimental conditions (2)





Extraction of EPO from clinical injection solution



Limit of detection of EPO from aqueous solution



LOD = 14.7 nM (S/N = 3 for the smallest peak)

Detection of EPO from spiked urine sample



Ultrafitration 1: Amicon Ultra-15 (MWCO 10 kD), to remove low molecular weight interfacing species;

Ultrafitration 2: Amicon Ultra-0.5 (MWCO 50 kD), to remove high molecular weight interfacing species, such as Tamm Horsfall glycoprotein (MW, 69 kD) and alpha-2-thiol proteinase inhibitor (MW, 72 kD).

Our proposed strategy



Outline

- Ø Nanoparticles-based extraction-capillary zone electrophoresis-native fluorescence detection of EPO
- Ø Boronate affinity monolithic capillaries for specific extraction of EPO

Why monolithic columns?



- Easy to prepare \rightarrow low cost
- Fast convective mass transfer \rightarrow high efficiency
- Open channel network \rightarrow low back pressure, fast separation

Boronate affinity



The cis-diol family:

Glycoproteins, glycopeptides, RNA, nucleosides, nucleotides, saccharides

Attractive features:

- Broad-spectrum affinity \rightarrow One ligand for all cis-diol biomolecules
- Covalent reaction \rightarrow High specificity
- Reversible \rightarrow Easy-to-control capture/release
- Fast desorption speed \rightarrow Nice peak shape, low carryover
- Eluted under acidic solution \rightarrow Compatible with MS

Boronate affinity monolithic capillaries with different features and functions

Name	monomer/crosslinker	characteristics	Specificity	Publication
Poly (VPBA- co-EDMA)	$H_{O}^{HO} \rightarrow H_{O}^{HO} + H_{O}^{O} \rightarrow H_{O}^{HO} + H_{O}^{O} \rightarrow H_{O}^{HO} + H_{O}^{O} \rightarrow H_{O}^{HO} + H_{O}^$	Hydrophobic, loading pH≥8.0	Poor	J Chromatogr A, 2009, 1216, 4768–4774
Poly (VPBA- co-MBAA)		Hydrophilic, loading pH ≥ 8.0	Good	J Chromatogr A, 2009, 1216, 8421–8425.
Poly (SPBA- co-MBAA)	$HO_{B} \rightarrow S_{O} \rightarrow H \rightarrow S_{O} \rightarrow H \rightarrow S_{O} \rightarrow S_{$	Hydrophilic, loading pH ≥ 7.0 Secondary separation capability	Good	Chem Commun, 2011, 47, 5067 - 5069.
Teamed boronate affinity	$H_{0} = 0^{H}$ $H_{1} = 1$ $H_{1} = 1$ $H_{1} = 1$ $H_{2} = 1$	Weakly hydrophobic, loading pH ≥ 7.0	Good	Angew Chem Int Ed, 2009, 48, 6704-6707.
Wulff-type boronate	OH H N-OH	loading pH ≥ 5.5	Good	Chem Commun, in press (DOI:10.1039/c1cc11096a).
Protein A/G like		loading pH ≥ 7.0, Highly selective to antibodies	Specific to antibodies	under preparation

Selective extraction of glycoproteins by G2 monolith at basic pH



(A) Direct MALDI-TOF MS analysis of a 1:1:1 mixture of the glycoprotein RNase B with the nonglycoproteins RNase A and myoglobin; (B) Direct MALDI-TOF MS analysis of the glycoprotein RNase B; (C) Extraction of RNase B from the mixture in (A) using the poly (VPBA-co-MBAA) and followed with MALDI-TOF MS analysis.

LOD: 10^{-11} mol RNase B (S/N = 200)

Improved sensitivity, selective!

Unpublished data

Specific extraction of EPO by G2 monolith at physiological pH



(A) Direct MALDI-TOF MS analysis of a 1:1:1 mixture of EPO, HRP and BSA; (B) MALDI-TOF MS analysis of EPO extracted from the mixture in A; (C) MALDI-TOF MS analysis of EPO extracted from EPO spiked human serum.

EPO: erythropoietin; HRP: Horseradish peroxidase; BSA: Bovine serum albumin

LOD: 10^{-10} mol EPO (S/N = 28)

Unpublished data

G6 monolith for antibody recognition, purification and immobilization





Most non-antibody glycoproteins tested were not retentive.



All antibodies tested were captured



Working at neutral pH A polymeric monolith exhibiting protein A/G-like affinity to antibody Sharp peaks

Identification of species extracted by G6 monolith from human serum



MALDI TOF MS of HSA + Anti-AFP

Specific extraction of EPO using Anti-EPO antibody immobilized monolith



MALDI TOF MS of EPO extracted by the anti-EPO antibody immobilized BA column from an aqueous EPO solution

Concluding remarks

- 1. A couple of new techniques have been developed for the extraction, separation and detection of EPO, including magnetic particles-based immuno-affinity extraction, monolith-based immuno-affinity extraction, capillary zone electrophoresis, and laser-induced native fluorescence. They provide useful tools for the analysis of samples with high EPO concentration and less complicated composition.
- 2. Due to the very limited concentration of EPO in real samples and the highly interfering matrix, anti-doping analysis of EPO is still a very challenging task. To address these issues, ultrasensitive detection and effective sample pretreatment approaches are highly desired.
- 3. Mass spectrometry-based proteomic analysis might be a useful alternative for anti-doping analysis of EPO, for which the boronate affinity monolithic capillaries might be a useful sample enrichment platform.

Acknowledgement

Funding agencies:

Ø World Anti-Doping Agency (WADA)
Ø The National Natural Science Foundation of China (NSFC)
Ø The Ministry of Science and Technology of China (MOST)



Thank all students in the lab!



Thank you for your attention!