

WADA <u>Technical Letter</u> – TL10

Document Number:	TL10	Version Number:	3.1
Written by:	WADA Science		
		Approved by:	WADA Executive Committee
Reviewed by:	WADA Laboratory Expert Group		
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IN SITU FORMATION OF EXOGENOUS COMPOUNDS IN URINE SAMPLES

1.0 Introduction

WADA wishes to draw the attention of the <u>Laboratories</u> to the following issues that may affect <u>Laboratory</u> operations. This pertains, in particular, to the various cases of formation of prohibited steroids or *Metabolites* originating from steroids normally present in urine *Samples* (*i.e.* non-prohibited sources) as a result of enzymatic (microbial) activities.

The impact of these metabolic *in situ* biotransformations may be different between the "A" and the "B" *Samples*. Depending on microbes' nature and growth, less frequently observed reactions could happen (*e.g.* formation of testosterone, Δ^1 -dehydrogenated steroids). Consequently, the formation of the 5 α - and 5 β -androstanediones (free form), the most common indicator of microbial modifications of the urinary steroids, is not always observed.

The greater sensitivity of the GC-MS/MS instruments permits the detection and confirmation of steroids that may have been formed by *in situ* enzymatic reactions. Therefore, <u>Laboratories</u> should be cautious when detecting low levels of steroids that could be formed microbially, particularly in the absence of their major *Metabolites*. In fact, screening for steroids (parent compounds) that are expected to be extensively metabolized following administration (*e.g.* nandrolone, androstenedione, boldione) should be carefully considered as this may cause incorrect interpretations and consequently erroneous decisions. For such *Prohibited Substances*, appropriate *Metabolites* (*e.g.* 19-NA for 19-norsteroids ^[11]) should be targeted for analysis. It should also be borne in mind that performing an enzymatic hydrolysis (especially overnight) directly on a urine *Sample* without a preliminary extraction, may exacerbate already present microbial activity and increases the risks of side-reactions.

The global pattern of *Metabolites* must always be evaluated by the <u>Laboratory</u>. For example, the presence of boldione without boldenone and its other main *Metabolites* should alert the <u>Laboratory</u> and trigger more investigations (*e.g.* verifying that the steroids are in the conjugated form and not free, performing GC/C/IRMS, when possible, in accordance with the TD IRMS ^[2]).

Examples of these biotransformations include:

- *i.* The formation of androst-4-ene-3,17-dione, 5α and 5β -androstanediones;
- *ii.* The formation of Δ^1 steroids such as boldenone, boldione and their *Metabolites*, androst-1-ene-3,17-diol from endogenous steroids ^[3, 4], as well as the formation of prednisone and prednisolone from endogenous cortisone and cortisol ^[5], respectively;
- *iii.* The formation of 19-norsteroids from demethylation of endogenous steroids (*e.g.* 19-NE from etiocholanolone, 19-NA from androsterone ^[6]);



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- *iv.* The formation of testosterone (free form, reported in some *Samples*; most frequently from female *Athletes*);
- *v.* The formation of reduced (5 α and 5 β -) *Metabolites* from 17 α -methyltestosterone when added as an internal standard (see TL08^[7]).

Excepting iii., where it has been shown that the conversion to 19-norsteroids occurs with glucuroconjugated steroids, the steroids produced will usually be found in the free form.

2.0 Analysis and Reporting Requirements

<u>Laboratories</u> should consider the following course of actions in the presence of steroid *Metabolites* that may have been formed by microbial activity:

- Check for signs of microbial activity [*e.g.* ratio of 5α -androstanedione (5α AND) to Androsterone (A), ratio of 5β -androstanedione (5β AND) to Etiocholanolone (Etio)] during the Initial Testing Procedure (ITP);
- During the <u>Confirmation Procedure</u> (<u>CP</u>), verify the presence of *Metabolites* in the free form instead of their expected conjugated state (*e.g.* testosterone in free form);
- Perform a <u>CP</u> using extraction prior to enzymatic hydrolysis to avoid inducing enzymatic conversions from microbes already present in the *Samples*. Do not add internal standards that may convert into the *Prohibited Substances*;
 - [Comment: In principle, it is recommended that <u>Laboratories</u> incorporate solid phase extraction (SPE) to clean up the Sample prior to the enzymatic hydrolysis in their chromatographic-mass spectrometric <u>CP</u>s. However, if the side products have already been formed prior to the enzymatic hydrolysis, **SPE will have no impact**.]

• Verify the presence of expected *Metabolites* according to known substance metabolic profiles.



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3.0 References

[1] *WADA Technical Document* TD NA: Harmonization of Analysis and Reporting of 19-Norsteroids related to Nandrolone.

[2] WADA Technical Document TD IRMS: Detection of Synthetic Forms of Prohibited Substances by GC/C/IRMS.

[3] Schänzer W., *et al.* Endogenous production and excretion of boldenone (17β-hydroxyandrosta- 1,4dien-3-one), an androgenic anabolic steroid, in: M. Donike, H. Geyer, A. Gotzmann, U. Mareck-Engelke (Eds.), *Recent Advances in Doping Analysis 2* Sport und Buch Strauß, Cologne, 1994; 211.

[4] Verheyden K., *et al.* Excretion of endogenous boldione in human urine: influence of phytosterol consumption. *J Steroid Biochem Mol Biol.* **117**(1-3): 8-14, 2009.

[5] Fidani M. et al. Presence of endogenous prednisolone in human urine. Steroids 78(2): 121-126, 2013.

[6] Grosse J *et al.* Formation of 19-norsteroids by *in situ* demethylations of endogenous steroids in stored urine samples. *Steroids* **70**: 499-506, 2005.

[7] WADA <u>Technical Letter</u> TL08: Use of Internal Standards.

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